

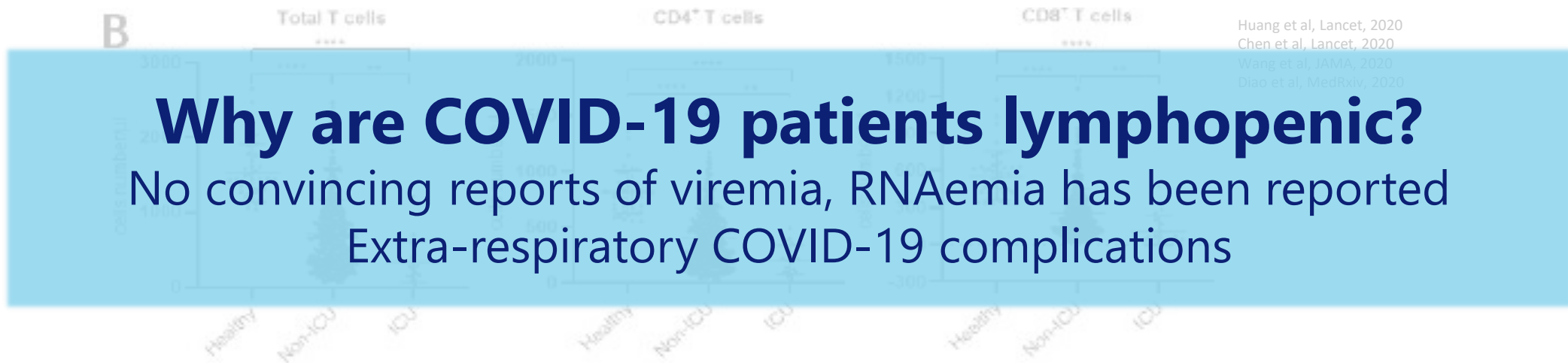
SARS-CoV-2-specific T-cell responses in COVID-19 patients

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Role for T-cells in COVID-19

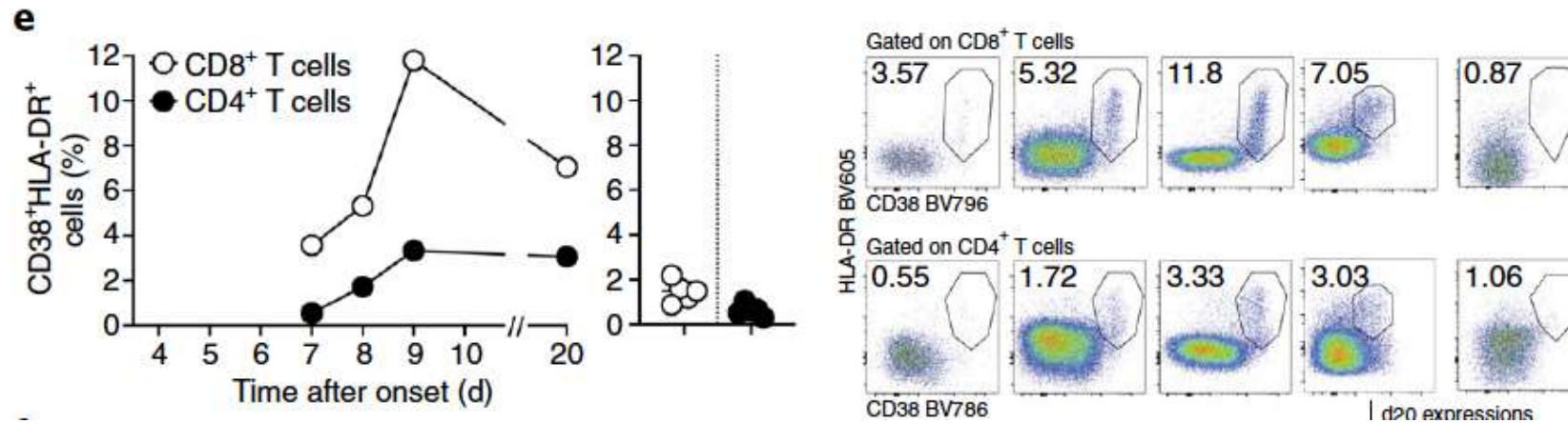
- COVID-19 patients present with low CD3⁺, CD4⁺ and CD8⁺ T cell counts



- Lymphopenia is associated with disease severity

T-cell activation in COVID-19

- Increase in activated T-cells in PBMC fraction during COVID-19



Thevarajan et al, Nat Med, 2020

SARS-CoV-2-specific T-cells

- What do we know about SARS-CoV-2-specific T-cells?

- ±10 published papers out, many more preprints

- Role of SARS-CoV-2-specific T-cells in disease is not yet understood

Lymphopenia and immune hyperresponsiveness
complex interaction between SARS-CoV-2 and the immune system

- CD4⁺ T-cell responses were correlated to positive outcomes in SARS-CoV-2 that is not fully understood

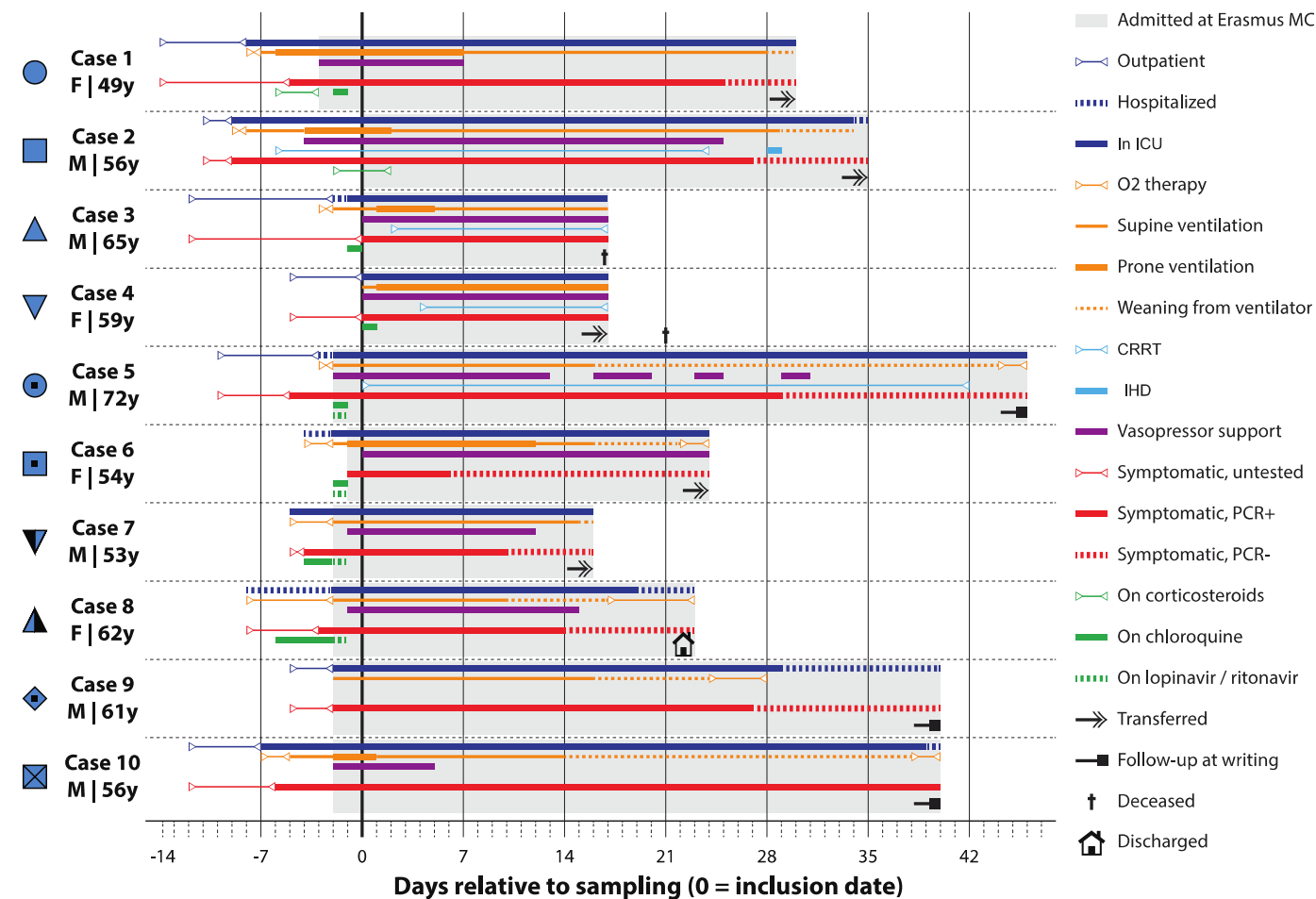
Channappanavar et al, Imm Res, 2014
Li et al, J Immunol, 2008

- Immune hyperreactivity and high levels of cytokines observed in severe COVID-19

- IL-6, IL-10, IP10, etc

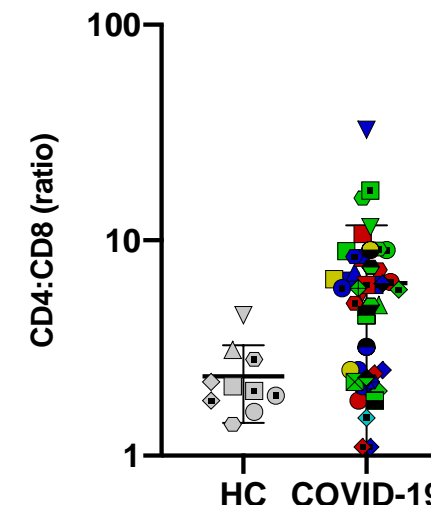
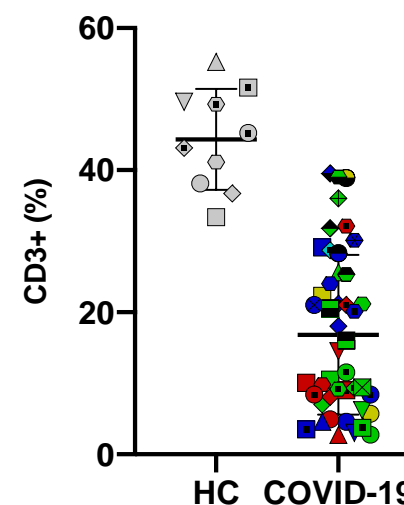
Zhou et al, Lancet, 2020
Evangelos et al, Cell, 2020

Study cohort (Acute Respiratory Distress Syndrome)

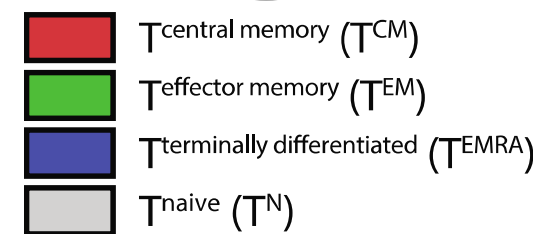
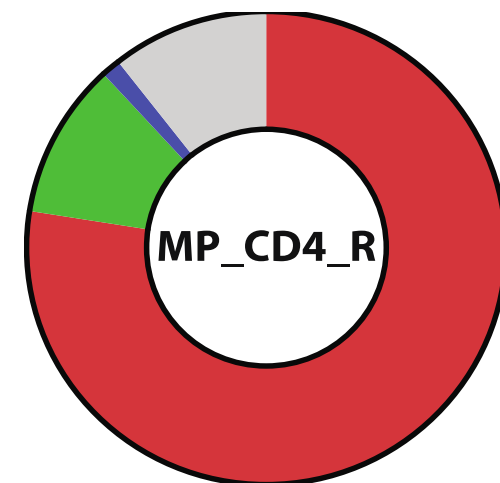
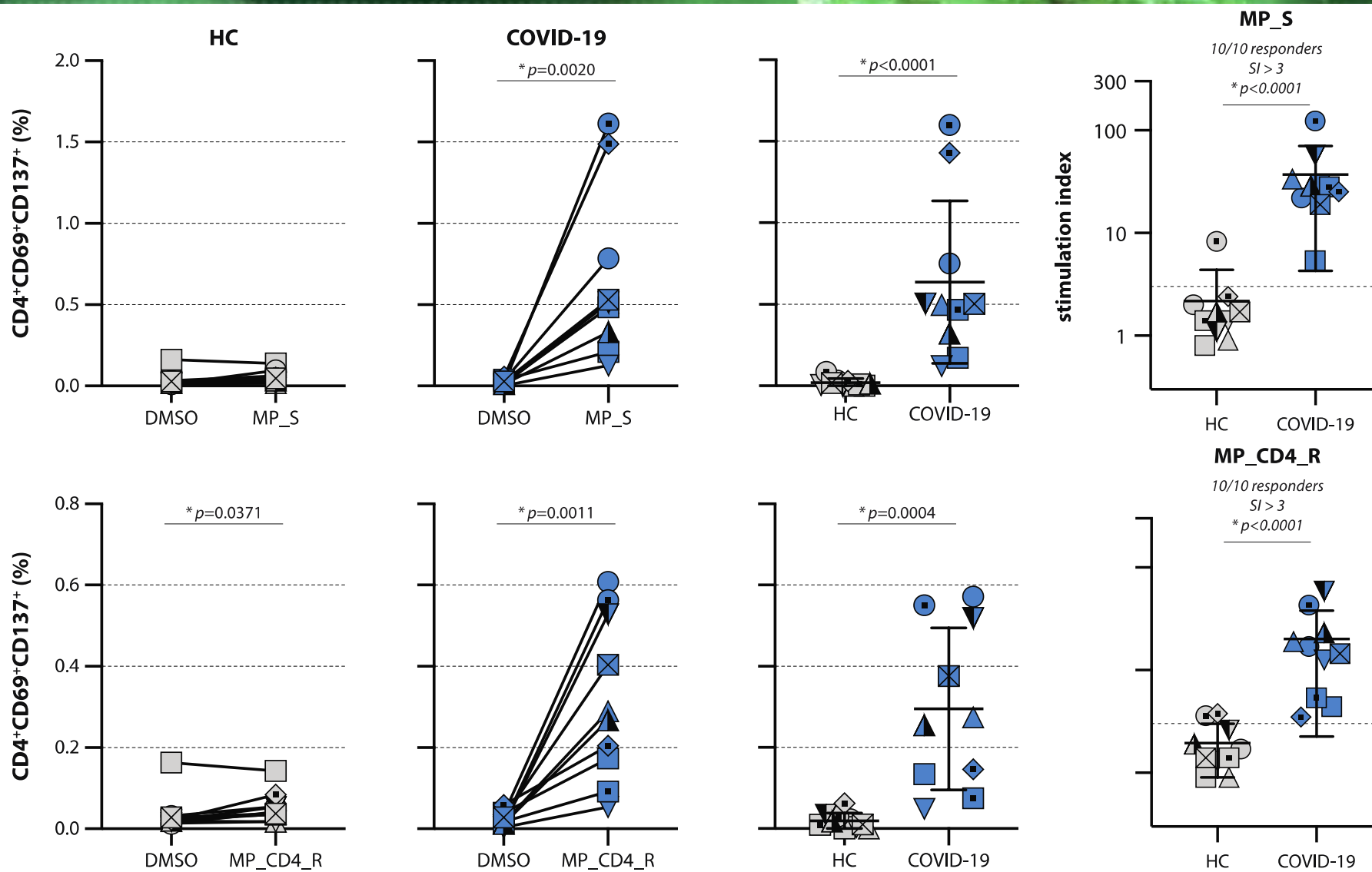


Study cohort

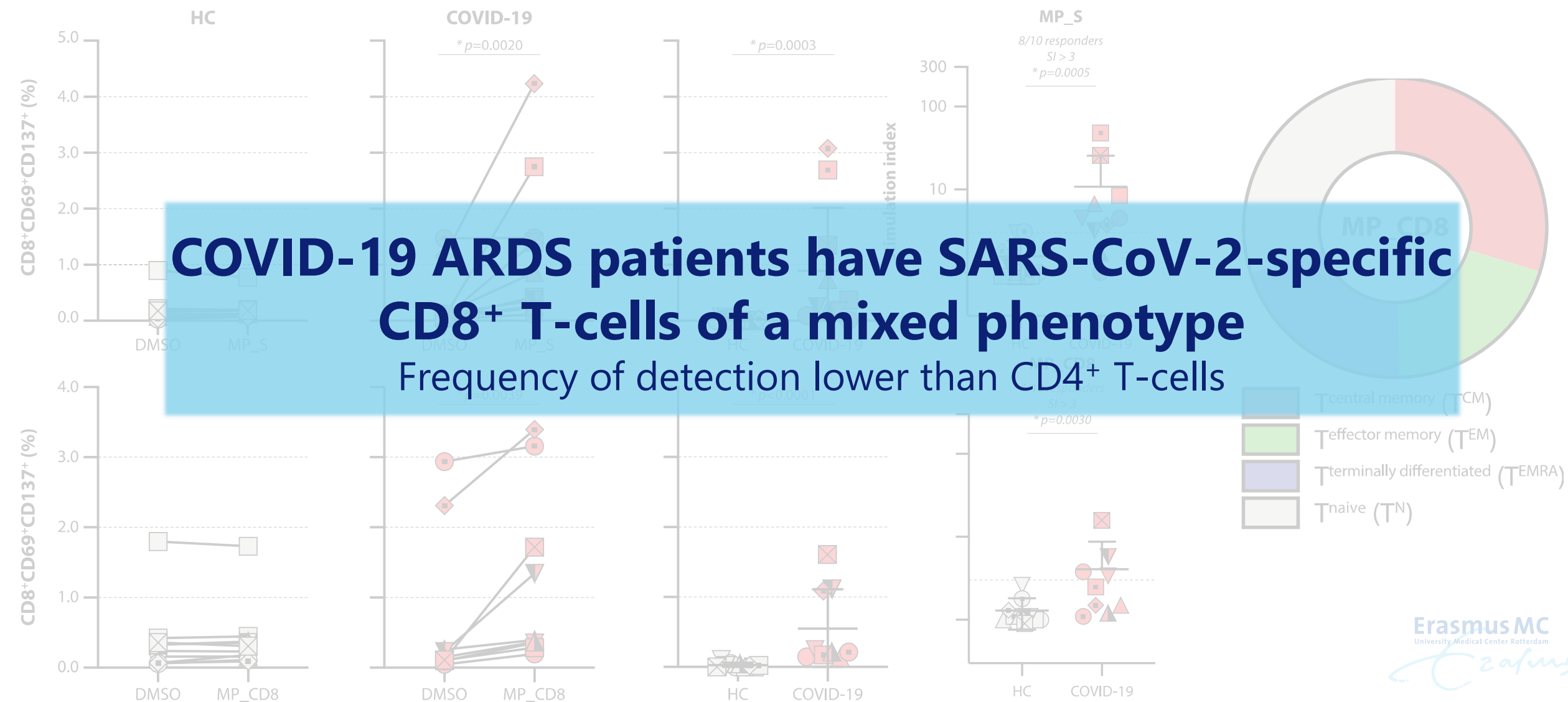
- N=10 COVID-19 ARDS patients (expanding)
- N=10 age-matched healthy controls



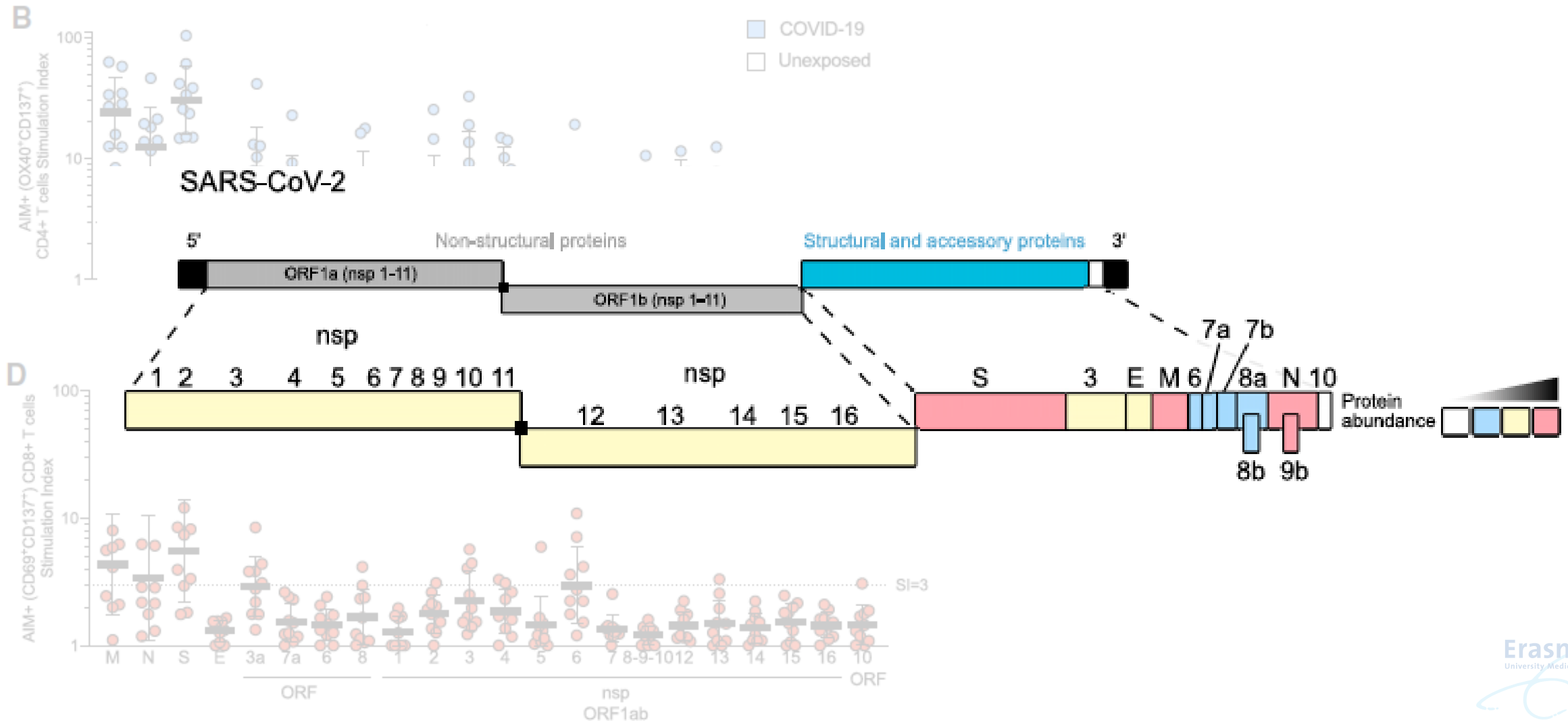
SARS-CoV-2-specific CD4⁺ T-cells



SARS-CoV-2-specific CD8⁺ T-cells



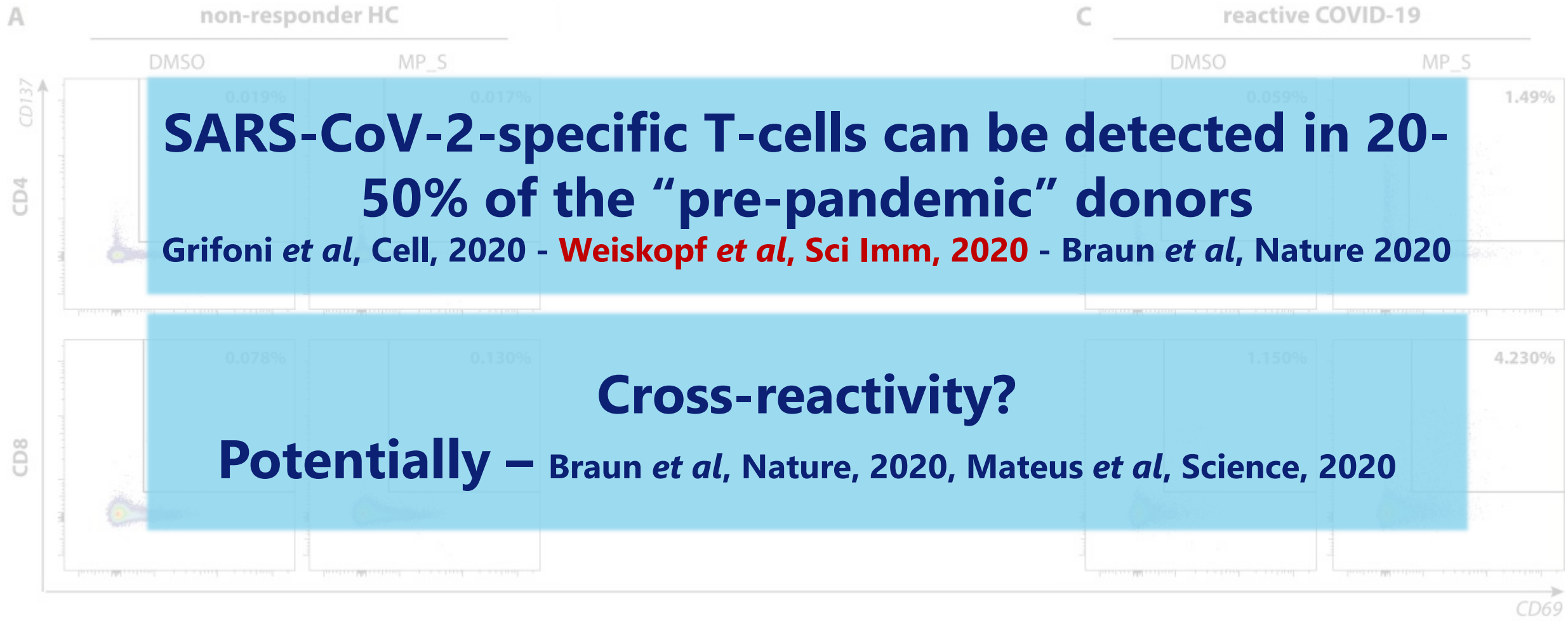
S, M and N are the prominent targets



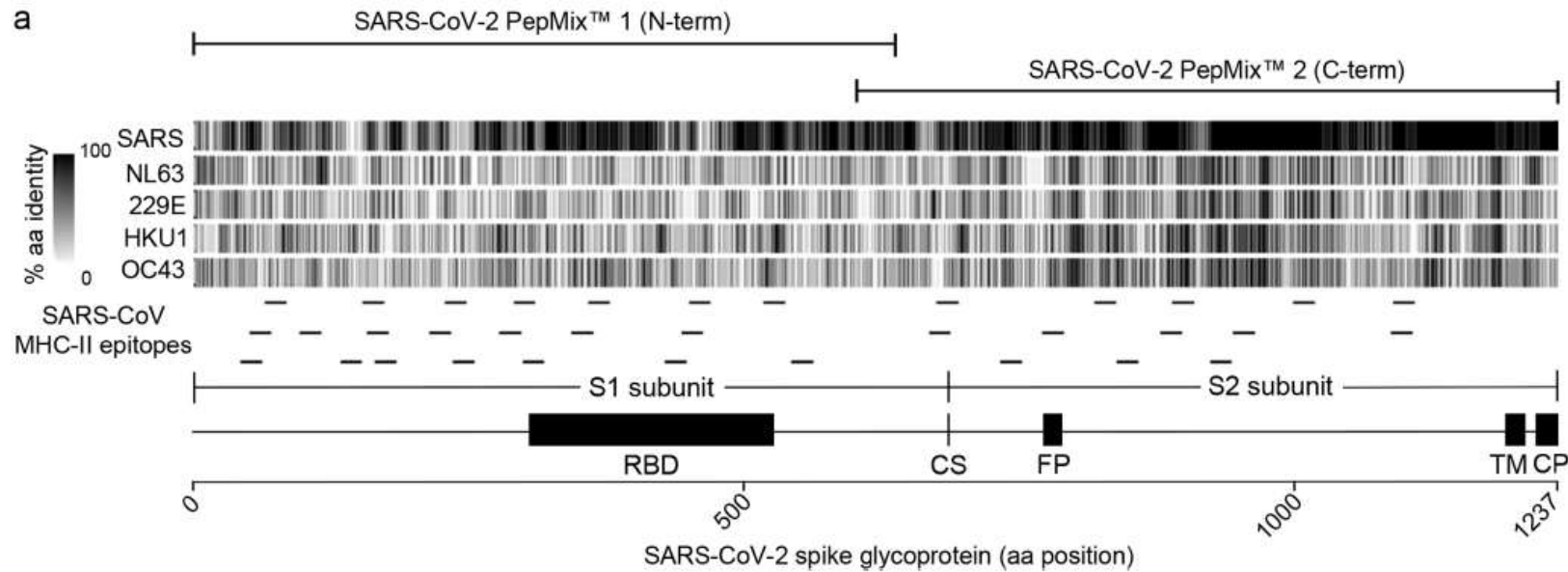
SARS-CoV-2-specific T-cells - OVERVIEW

		CD4 ⁺ MP_S	CD4 ⁺ MP_CD4_R	CD8 ⁺ MP_S	CD8 ⁺ MP_CD8
Weiskopf et al, Sci Imm, 2020	HC	1/10 (10%)	2/10 (20%)	1/10 (10%)	0/10 (0%)
	COVID-19	40/45 (89%)	30/42 (71%)	27/45 (60%)	15/41 (37%)
Braun et al, Nature, 2020	HC	24/68 (35%)			
	COVID-19	15/18 (83%)			
Grifoni et al, Cell, 2020	HC	1/11 (9%)	4/11 (36%)		1/10 (10%)
	COVID-19	10/10 (100%)	10/10 (100%)		7/10 (70%)

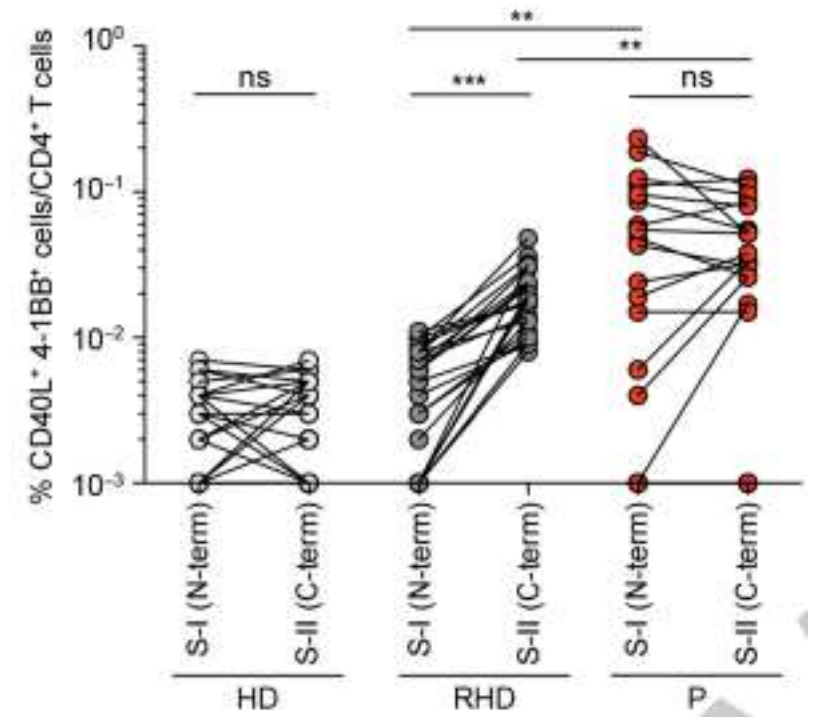
Cross-reactive T-cells in HC



Cross-reactive T-cells in HC

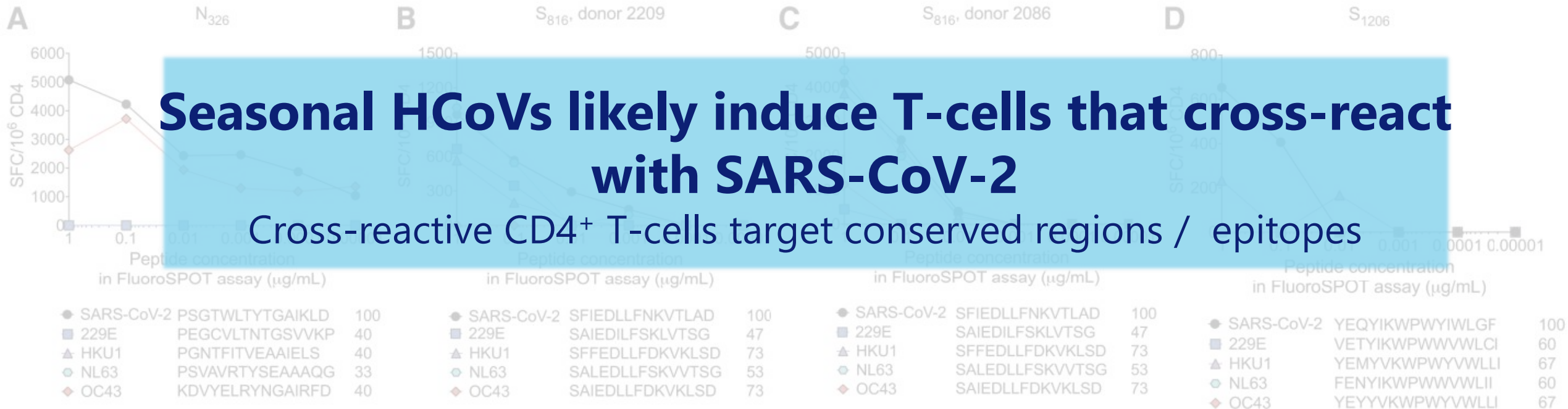


Braun et al, Nature, 2020



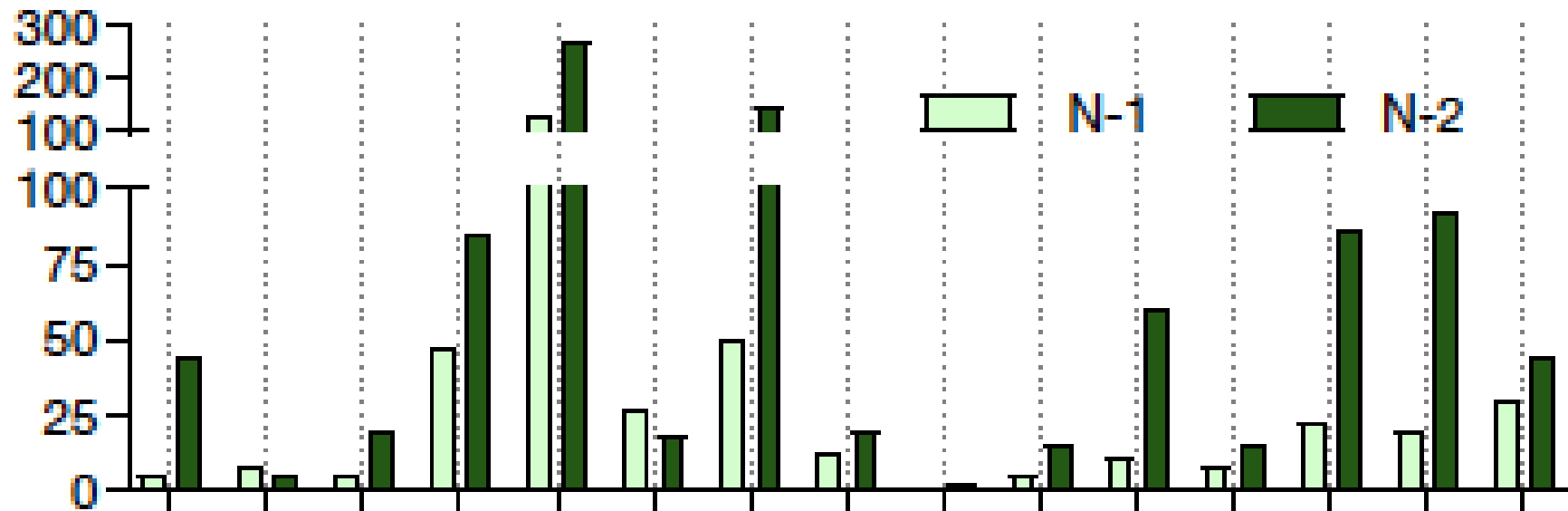
Cross-reactive T-cells in HC

- T-cell lines generated after peptide stimulation of PBMC from pre-pandemic donors



SARS-CoV-specific T-cells are long-lived

- PBMC isolated from SARS recovered individuals 17 years after disease
 - Stimulated with N peptide pool



Le bert et al, Nature, 2020

Summary and Discussion

- SARS-CoV-2-specific CD4⁺ and CD8⁺ T-cells detected in blood of COVID-19 patients
 - S, M and N are the dominant targets

- (Dominant effector and Th1) cytokine production in response to viral antigen

**What is the role of (cross-reactive) T-cells
in
(prevention of) disease??**

- Reactive T cells were detected in HC after MP stimulation → cross-reactivity

- Also reported by Grifoni *et al* (US), Braun *et al* (Germany), Le Bert *et al* (Singapore) and Meckiff *et al* (UK)
 - Possible induction by circulating seasonal common cold coronaviruses (NL63, HKU1)

- SARS-CoV-specific T-cells are long-lived

- Detection of SARS-CoV-2-specific T-cells is an accurate measure of exposure

Erasmus MC, Viroscience

- Katharina Schmitz
- Matthijs Raadsen
- Nisreen Okba
- Richard Molenkamp
- Marion Koopmans
- Bart Haagmans
- Eric van Gorp
- Rik de Swart



Erasmus MC, ICU

- Henrik Endeman
- Johannes vd Akker



LJI, San Diego, USA

- Daniela Weiskopf
- Alba Grifoni
- Alessandro Sette



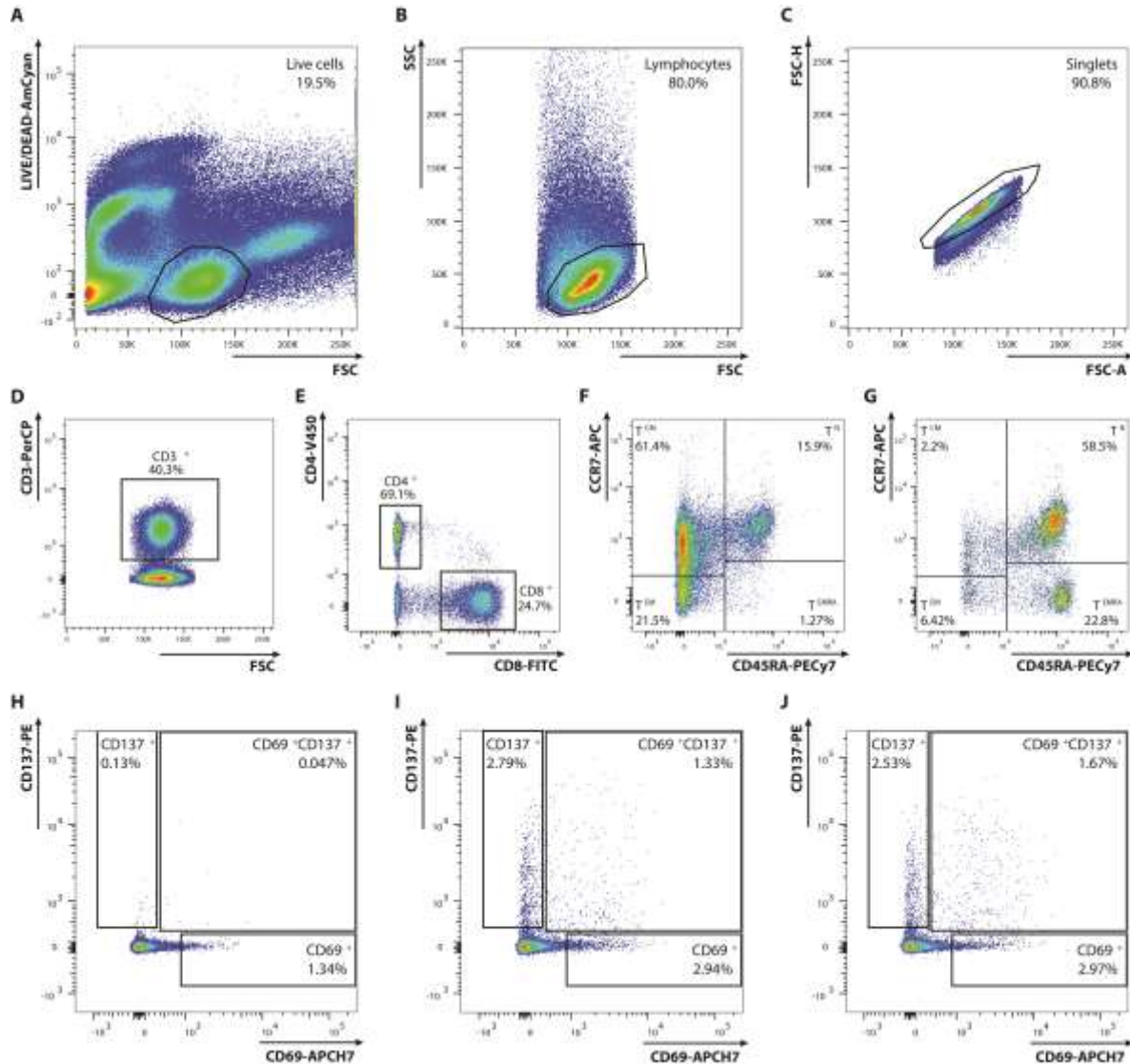
MegaPool (MP) stimulations

- **Method:** stimulate PBMC with peptide pools, detect T cell activation
- **Specific peptide pools used (MegaPools, MPs)**
 - MPs are pools with large numbers of peptides generated by sequential pooling and lyophilization.
 - Can incorporate **overlapping peptides**, or **predicted** or **experimentally validated epitopes**

Activation-Induced Markers (AIM)

- **Method:** stimulate PBMC with peptide pools, detect T cell activation
 - Upregulation of activation markers **CD69** and **CD137** (20h stimulation)
 - Intracellular detection of IFN γ and TNF α (8h stimulation)
 - Cytokines measured in culture sup with multiplex beads assay (20h stimulation)

Activation-Induced Markers (AIM)



■ Gating strategy:

- A: LIVE cells
- B: Lymphocytes FSC / SSC
- C: Single cells
- D: CD3⁺ cells
- E: CD4⁺ / CD8⁺ cells
- F/G: Memory phenotyping on basis of CD45RA / CCR7
- H: CD69 vs CD137 **DMSO (- control)**
- I: CD69 vs CD137 **CMV (+ control)**
- J: CD69 vs CD137 **MP_S**

■ Double positive cells in J used in graphs

T-cells in absence of seroconversion

Figure 1

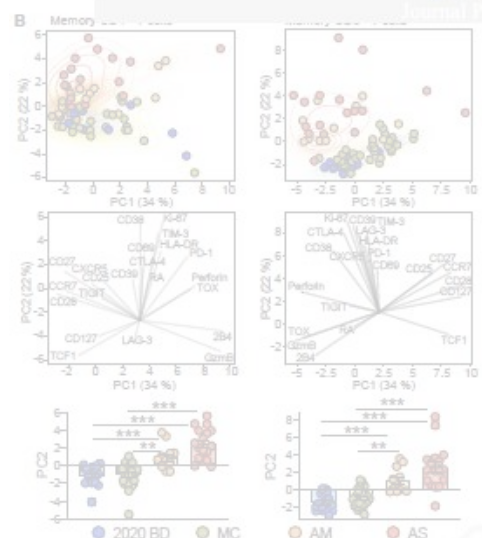


Figure 2



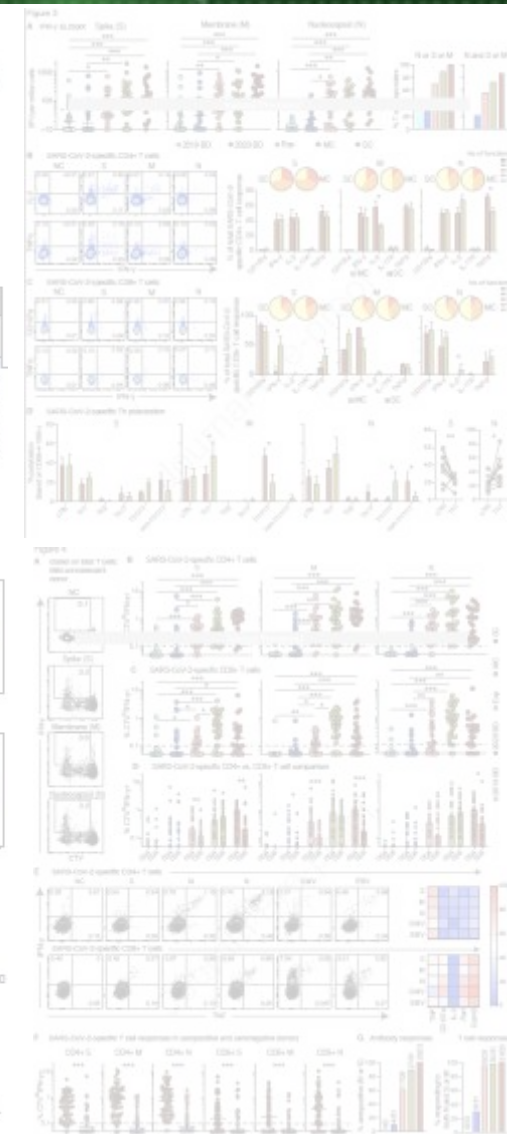
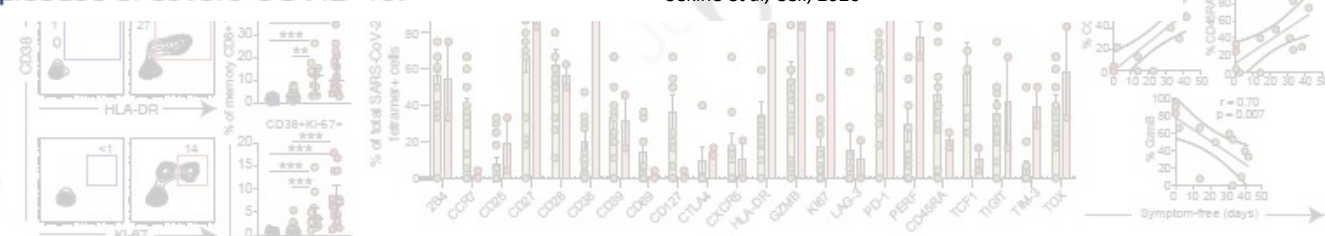
Figure 2B: Dot plots showing the expression of various markers on EBV-specific CD8+ T cells.



SUMMARY

SARS-CoV-2-specific memory T cells will likely prove critical for long-term immune protection against COVID-19. We here systematically mapped the functional and phenotypic landscape of SARS-CoV-2-specific T cell responses in unexposed individuals, exposed family members, and individuals with acute or convalescent COVID-19. Acute phase SARS-CoV-2-specific T cells displayed a highly activated cytotoxic phenotype that correlated with various clinical markers of disease severity, whereas convalescent phase SARS-CoV-2-specific T cells were polyfunctional and displayed a stem-like memory phenotype. **Importantly, SARS-CoV-2-specific T cells were detectable in antibody-seronegative exposed family members and convalescent individuals with a history of asymptomatic and mild COVID-19.** Our collective dataset shows that SARS-CoV-2 elicits robust, broad and highly functional memory T cell responses, suggesting that natural exposure or infection may prevent recurrent episodes of severe COVID-19.

Sekine et al, Cell, 2020



Summary and Discussion



The Netherlands Organisation for Health Research and Development
Programme: Off Road 2016-2017

Project summary

Coronaviruses are endemic in humans and often cause a self-limiting infection with common cold-like symptoms. Some of the zoonotic high-threat emerging viruses are also coronaviruses: MERS-coronavirus (CoV) has recently been identified as novel zoonotic agent and is continuing to spill over to humans. Virus-specific cytotoxic T-cells, considered crucial for viral clearance, have not yet been detected for human coronaviruses. **The overall aim of this proposal is to detect T-cells specific for endemic- and MERS-CoV and determine whether these T-cells can be cross-reactive.** This will be addressed by probing blood samples from unique MERS-CoV-infected patients from the Middle East for endemic coronavirus-specific, MERS-CoV-specific and cross-reactive T-cells. As a proof of principle, I will develop a mouse model to prove that endemic coronaviruses can protect from MERS-CoV-related disease.